

# **An Open Platform, High Speed CD4 T- Cell Monitoring Method Using Immunogold Conjugates: A Method Aimed at Reducing Loss to Follow-Up in Resource-Poor Settings**

Peter Hansen<sup>1</sup>, David Dombkowski<sup>2</sup>, Michelle DeLelys<sup>2</sup>,  
Amanda Restell<sup>3</sup>, Dorothy Sylvia<sup>3</sup>, Donald Barry<sup>3</sup>, Sonia Kumar<sup>3</sup>,  
Artem Zeygerman<sup>3</sup>, Petra Krauledat<sup>3</sup>, Frederic Preffer<sup>2</sup>

<sup>1</sup>Union Biometrica Tech, Inc., Canaan, New York, USA

<sup>2</sup>Massachusetts General Hospital, Dept. of Pathology, Boston, MA, USA

<sup>3</sup>PointCare Technologies, Inc., Marlborough, MA, USA

# Background

## Standard CD4 T-cell Test:

- Costly instrumentation using lengthy immunofluorescence labeling methods
- Requires cold-chain shipping (maintain 2-8°C during shipping)
- Reagents need to be stored at 2-8°C
- Results are not usually available the same day

## Resource-Poor Settings:

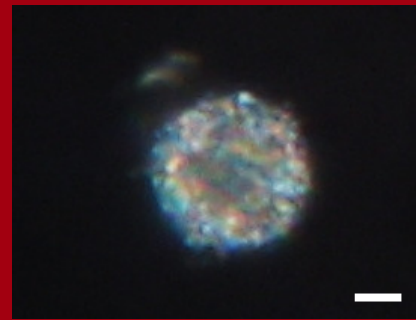
- Requires cost effective CD4 test method
- Unreliable power sources for refrigerated storage
- Patients distant from clinics often do not return for follow-up visits

# Methods

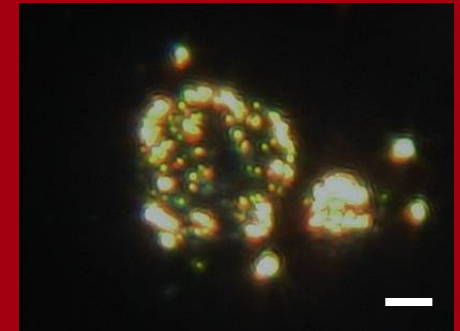
## Immunogold CD4 T-Cell Assay

- Minimal sample preparation can be completed in  $< 1$  min
- Cationic surfactant neutralizes surface charge of immunogold resulting in ultra-fast labeling, in as little as 45 seconds
- Reagents are temperature stable and do not require cold chain shipping or refrigerated storage
- CD4 clusters can be detected with relatively simple, cost effective optics

CD4-Negative Lymphocyte



CD4-Positive Lymphocyte Labeled with Immunogold

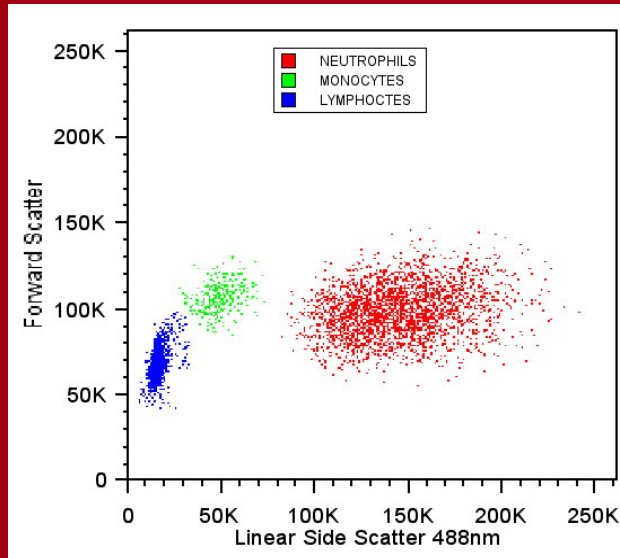


Darkfield reflectance images of lysed whole blood, scale bar is  $\sim 2\mu\text{m}$

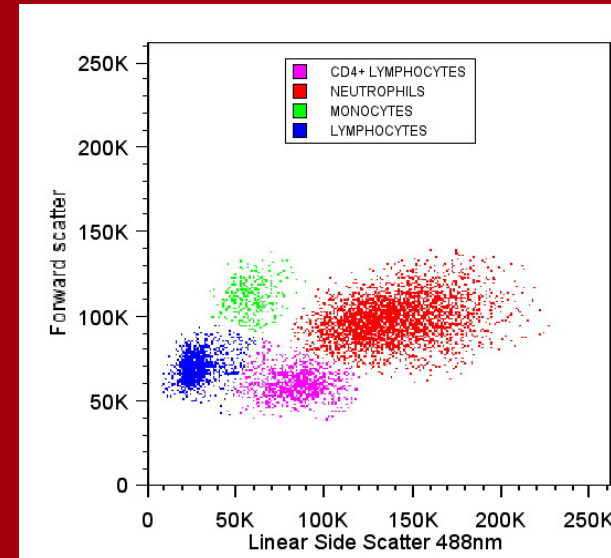
Reflectance images show dramatic increase in scattering for CD4+ cells labeled with targeted immunogold contrast agent

# Results

Unlabeled Light Scatter Plot



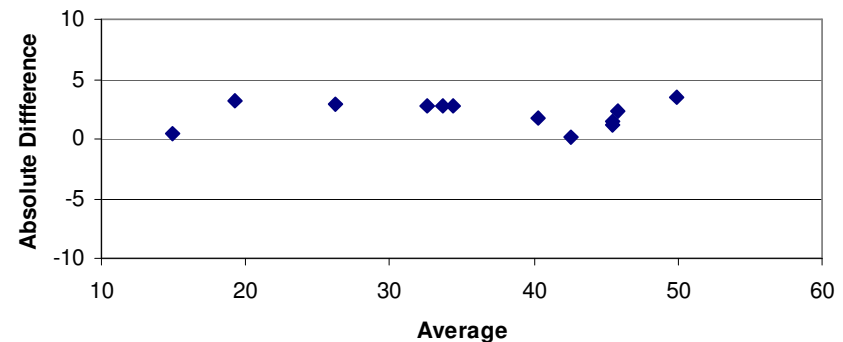
Immunogold Labeled Light Scatter Plot



Bland-Altman comparison of 12 samples (6 clinical, 5 normal, and 1 manipulated to a low CD4% of 15) run on the same flow cytometer using the standard fluorescence method vs immunogold assay shows excellent agreement between the two methods.

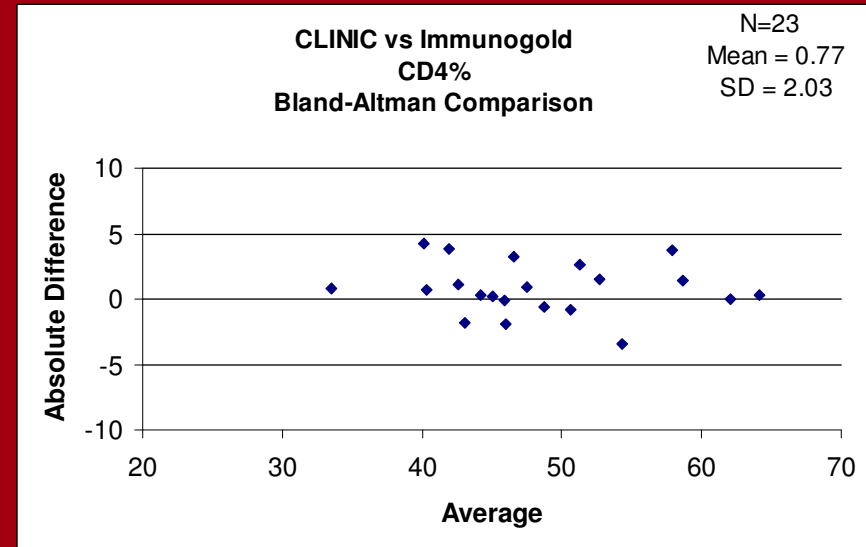
Fluorescence vs Immunogold  
CD4% Same Platform  
Bland-Altman Comparison

N = 12  
Mean = 2.1  
SD = 1.1



# Results

A Bland-Altman Comparison of 23 normal samples compares CD4% results from a flow cytometer using the immunogold assay to CD4% results obtained from a reference laboratory using the standard fluorescence analysis.



# Conclusions

- Bland-Altman shows good agreement between the standard fluorescence method and the immunogold light scatter method
- Currently deployed flow cytometers can use immunogold to identify CD4 T-cells
- Immunogold delivers immediate CD4 results
- Immunogold is unusually heat stable (42C continuously for one year) eliminating cold-chain shipping and refrigeration